

A quantitative comparison of microfossil extraction methods from ice cores

SANDRA O. BRUGGER,^{1,2} E. GOBET,^{1,2} F. R. SCHANZ,¹ O. HEIRI,^{1,2} C. SCHWÖRER,^{1,2}
M. SIGL,^{2,3} M. SCHWIKOWSKI,^{2,3,4} W. TINNER^{1,2}

¹Institute of Plant Sciences, University of Bern, Switzerland

²Oeschger Center for Climate Change Research, University of Bern, Switzerland

³Paul Scherrer Institute, Villigen, Switzerland

⁴Department for Chemistry and Biochemistry, University of Bern, Switzerland

Correspondence: Sandra O. Brugger <sandra.bruegger@ips.unibe.ch>

ABSTRACT. Microfossil records from ice archives allow vegetation, fire and land-use activity reconstructions on broad spatial scales. Samples typically contain low microfossil concentrations. Therefore, large ice volumes are often needed for palynology. Hence, it is crucial to extract maximum microfossil numbers through appropriate physical-chemical treatments. We compare six methods covering the main water reduction procedures: evaporation, filtration and centrifugation with snow samples. Adding a known number of *Lycopodium* marker spores prior to sample treatment and a second marker (*Eucalyptus*) after laboratory processing allows a quantitative microfossil loss assessment during pollen extraction. We applied the best-performing method (average loss of 22%) to high-alpine firn cores from Colle Gnifetti glacier for validation with a natural archive containing extremely low microfossil concentrations. We conclude that samples processed with different microfossil extraction protocols may give different results for pollen concentrations, percentages and ratios between different pollen types, especially if vesiculate conifer pollen is an important pollen assemblage component. We recommend a new evaporation-based method which delivers the smallest and least variable losses among the tested approaches. Since microfossil losses are inevitable during laboratory procedure, adding markers prior to sample processing is mandatory to achieve reliable microfossil concentration and influx estimates.

KEYWORDS: ice biology, paleoclimate, *Eucalyptus* marker, glacier, *Lycopodium* marker, palynology, pollen

1. INTRODUCTION

Only a handful of microfossil records from ice cores and surface snow samples are available at present (overview in Table 1), probably because the records are difficult to retrieve and the concentrations of the target material are low. In contrast to the more traditional archives of palynology (e.g. lakes and peat bogs), ice archives have specific advantages. For instance, they are well suited to address vegetation dynamics and land-use activities at subcontinental scales (Liu and others, 1998), since drilling sites on high-alpine glaciers are remote from microfossil sources and undesired local biases are absent. Further, they do not suffer from fine-scale disturbances that may affect lakes or peatlands such as soil erosion and related reworking issues. Ice cores usually provide excellent chronologies and high temporal resolution records, especially for the most recent 200 years where age-depth models can rely on absolutely-dated reference horizons (e.g. volcanic layers) and annual layer counting (e.g. Preunkert and others, 2003; Olivier and others, 2006; Jenk and others, 2009; Sigl and others, 2009; Herren and others, 2013; Konrad and others, 2013). Multiproxy climate and environmental evidence from the same cores (e.g. temperature reconstructions or chemical tracers for environmental variables; Eichler and others, 2011) contribute to assess past ecosystem dynamics.

Glaciers contain extremely low microfossil concentrations compared with lake and mire sediments, hence large ice

volumes are needed for quantitative microfossil analyses. Furthermore, given the difficulty of accessing the often remote drilling sites, ice core material is usually very limited and therefore it is crucial to extract a maximum of microfossils for analysis. This provides a major challenge for sample processing. Different methods have been used in the past to concentrate and extract pollen from ice or snow samples with three main water elimination approaches described in the literature: evaporation, centrifuging followed by decanting of the supernatant liquid and filtering methods (see Table 1 for an overview of the available methods). No qualitative or quantitative comparison of the different extraction methods is available so far. Differences among the approaches may impede comparison of microfossil ice core results, but this potential source of uncertainty is unexplored. Thus, the situation is very different than for pollen analysis in lake and mire sediments with well-recognized preparation procedures and protocols (Faegri and Iversen, 1989; Moore and others, 1991; Lang, 1994).

Marker spike application is a long-established and reliable method to estimate microfossil concentrations and influx (Benninghoff, 1962; Stockmarr, 1971; Peck, 1974; Birks and Birks, 1980; Maher, 1981; Birks and Gordon, 1985; Moore and others, 1991). Its use additionally allows to check for microfossil losses during sample preparation (Stockmarr, 1971). Most of the previous microfossil records

Table 1. Methods for microfossil extraction from ice, grouped according to the main water elimination procedure with an indication of marker use**Centrifugation**

Eichler and others (2011)* (EICHLER method)

Festi and others (2015)[†] (FESTI method)Vareschi (1934)[‡]; Ambach and others (1966)[‡]Fredskild and Wagner (1974)[‡]Feurdean and others (2011)[‡]**Evaporation**

Liu and others (1998; 2005; 2007)* (LIU method),

Reese and Liu (2002; 2005)*,

Reese and others (2003)*

Yang and others (2008)* (YANG method)

Filtration*Dissolving filter*Andreev and others (1997)[‡]

Hicks and Isaksson (2006)*

Koerner and others (1988)[‡]McAndrews (1984)[‡]

Short and Holdsworth (1985)* (SHORT method)

*Rinsing filter*Papina and others (2013)[‡]*Counting on filter*Nakazawa and others (2004, 2005, 2006, 2011)[‡]Uetake and others (2006)[‡]Santibañez and others (2008)[‡]

Bourgeois (1990; 2000), Bourgeois and others (2000; 2001)*

Approaches used for the method comparison in this paper with method-name in brackets. Filtration-based approaches are divided in methods that chemically dissolve the filter, rinse the microfossils with water from the filter surface or methods where microfossils are directly counted on the filter.

* Marker added, [†]No marker added, [‡]No information available.

from ice cores rely on marker application (e.g. Liu and others, 1998; Eichler and others, 2011; see Table 1 for an extensive list). Exceptionally, its use has been neglected in recent ice studies (Festi and others, 2015) because the utility of this standard palynological approach has been questioned (Festi and others, 2016). Here we address existing knowledge gaps and open questions in respect to the effects of sample preparation methods on pollen assemblages from ice core records with the following aims: (1) to compare different palynological extraction methods for snow, firn and ice core records; (2) to assess the risk of microfossil losses during the extraction procedure and thus the utility of markers in ice core samples; and (3) to propose an improved extraction protocol to gain reliable, accurate and comparable microfossil results from snow and ice samples with a minimum of loss during extraction.

2. MATERIALS AND METHODS

2.1. Marker tablets

To quantify the losses during the different extraction procedures, we added two different markers to the samples, one before and another after the physical and chemical treatments. We used *Lycopodium clavatum* (Batch number 3862 with 9666 spores per tablet \pm 671 Std dev.) and *Eucalyptus* marker tablets (Batch number 106720 with 13500 pollen grains \pm 210) provided by University of Lund (Maher, 1981). Before proceeding with microfossil extraction, we tested the reliability of the marker tablets by mixing one tablet of *Lycopodium* marker and one tablet of *Eucalyptus* marker. Ten of these marker tablet pairs were dissolved in 10% HCl following standard procedures for palynology (e.g. Moore and others, 1991). The marker suspensions were mounted on microscopic slides. We counted a sum of 1000 grains (*Lycopodium* + *Eucalyptus*) to estimate the accuracy of the expected marker ratio between *Lycopodium* and *Eucalyptus* grains.

2.2. Microfossil extraction methods

2.2.1. Snow replicate samples

To compare the different microfossil extraction methods from snow and ice, we collected 10 surface snow samples (uppermost 30 cm) of ~4 kg at Jungfraujoch (Swiss Alps, 46° 32' 54.6" N, 7° 58' 58.8" E, 3400 m a.s.l.) in July 2016 after conifers flowered in late spring (Lauber and Wagner, 2012). The 10 samples (Jung-1 – Jung-10) were transported frozen to the laboratory and separated in small pieces in the freezing chamber at –17°C to avoid melting. The pieces were homogenized well before dividing them into six replicate subsamples, resulting in a total of 60 test samples. Each of the six replicate samples weighed between 370 and 400 g (corresponding to a volume of 370–400 mL). The microfossil extraction followed five methods from the literature covering the main physical water reduction procedures of evaporation (Liu and others, 1998; Yang and others, 2008), centrifuging and decanting (Eichler and others, 2011; Festi and others, 2015), and filtration (Short and Holdsworth, 1985; Fig. 1), followed by a chemical treatment involving several centrifuging steps. For simplicity, we labeled the applied methods using the first author's name in the publications (e.g. LIU method for the method described in Liu and others, 1998; and accordingly, YANG, EICHLER, FESTI and SHORT method; Table 1). The evaporation-based methods LIU and YANG differ in a HF treatment included in the latter method, which requires additional steps for the laboratory protocol. The EICHLER and FESTI methods follow a comparable protocol except for the addition of *Lycopodium* tablets and an alcohol treatment to lower the water surface tension in the samples used by FESTI. Microfossil concentration methods based on filtration, with direct counts of microfossils on the filter, were not considered in this study due to the strong limitations in the taxonomic resolution that can be achieved (Nakazawa and others, 2005).

We tested a new extraction method, which we refer to as BRUGGER method (Fig. 1). This method starts with evaporation in a drying cabinet (1 week at 70°C for 400 mL sample volume) to reduce the water content to ~20 mL. The samples are then transferred to 50 mL tubes including

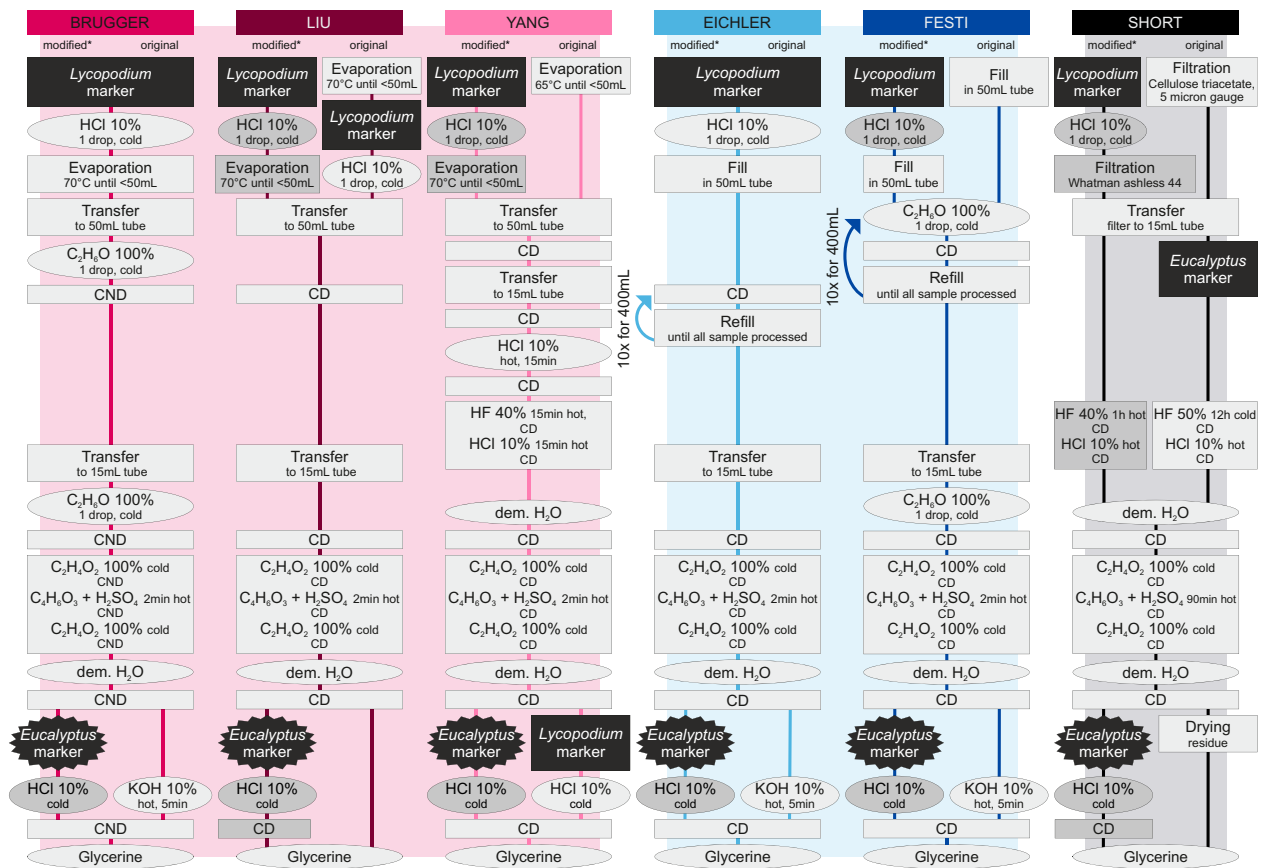


Fig. 1. Flowcharts for ice and snow sample extraction methods: BRUGGER = this study, LIU = Liu and others (1998), YANG = Yang and others (2008), EICHLER = Eichler and others (2011), FESTI = Festi and others (2015), SHORT = Short and Holdsworth (1985). Right column for each method indicates original method description. Left column (shaded in dark grey) indicates required steps to add a second marker (*Eucalyptus*) and the deviation from the original protocol for the standardized method comparison of this paper with two markers. CND = centrifuging, shock-freezing in liquid nitrogen, decanting, CD = centrifuging, decanting. C_2H_6O = ethanol, $C_2H_4O_2$ = glacial acetic acid, $C_2H_6O_3$ = acetic anhydride, H_2SO_4 = sulphuric acid, dem. H_2O = demineralized water.

careful rinsing of the original containers. After each centrifugation step, we shock-freeze the bottom liquid of the tubes, which contains the sunken microfossils, with liquid nitrogen and decant the remaining water. Subsequently, the samples are transferred to 15 mL tubes. Acetolysis and a 10% KOH treatment follow before mounting the samples in glycerine (for details on acetolysis see Moore and others, 1991). The protocol is comparable with the method described in Liu and others (1998) but it includes the step of shock-freezing the bottom liquid after each centrifugation step to avoid material losses. Additionally, we add a drop of ethanol (C_2H_6O) to the centrifuge tubes before centrifuging for all protocol steps with water to reduce floating of microfossils due to water surface tension effects (Dietrich, 1923). Finally, to test for microfossil losses during the physical and chemical procedure we added one *Lycopodium* tablet to each snow replicate sample prior to sample processing (Stockmarr, 1971). After sample processing, we added one *Eucalyptus* marker tablet and we applied once more 10% HCl to dissolve the tablet before the last centrifugation step (Fig. 1). *Lycopodium* spores and *Eucalyptus* pollen are nonvesiculate grains and have a diameter of ~25 and 20 μm , respectively.

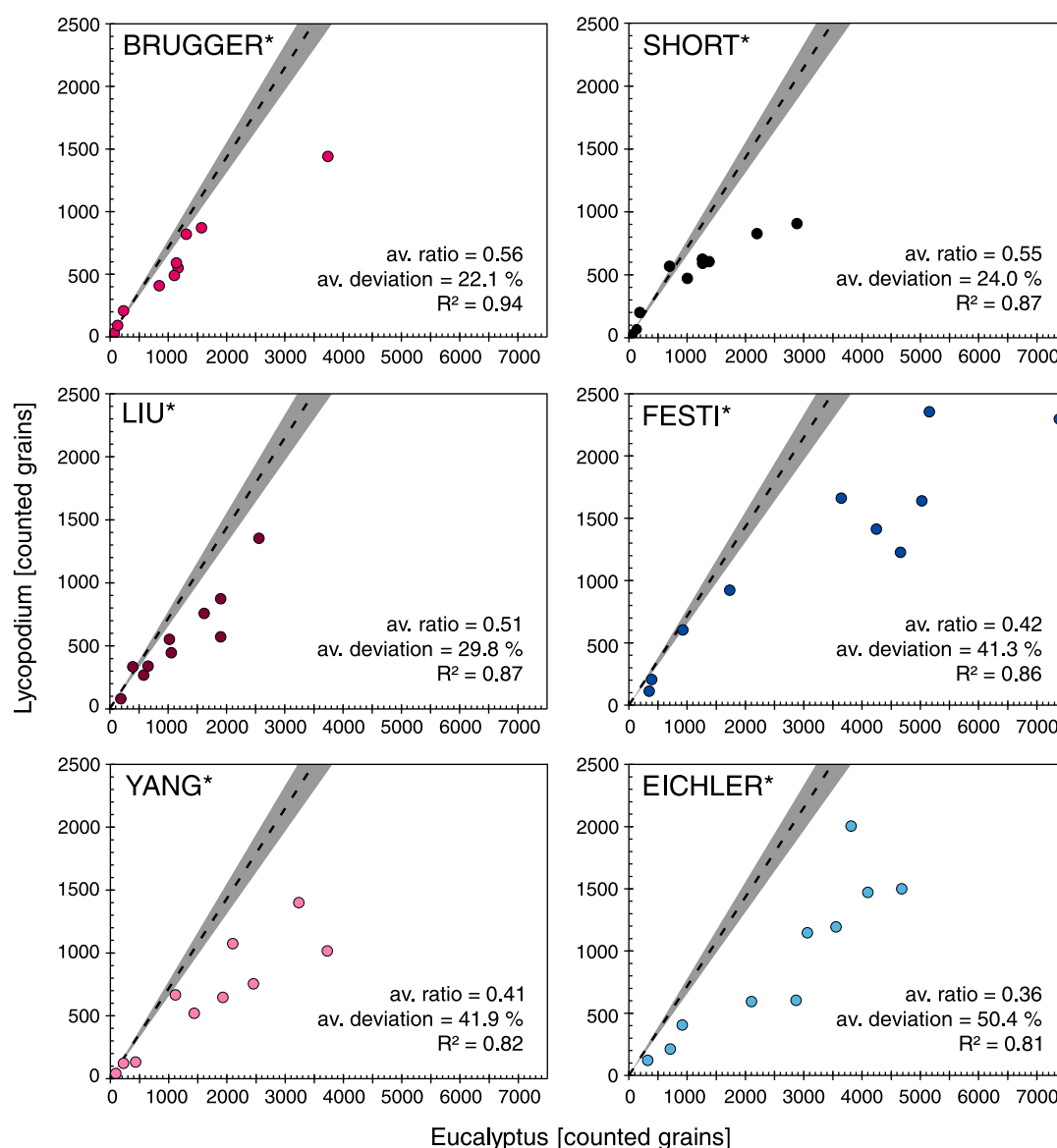
2.2.2. High-alpine ice core samples

To further evaluate our new method, we applied it to high-alpine ice core samples from Colle Gnifetti glacier. This

glacier saddle forms part of the Monte Rosa massive in the Swiss Alps (45°55'50"N, 7°52'33"E, 4450 m a.s.l.). The ice core was drilled in September 2015 and stored frozen at the Paul Scherrer Institute in Villigen. In total 18 samples from adjacent core segments of varying length (Samples Colle-1 – Colle-18) spanning 2015–01 AD were cut in the freezing chamber. We included one additional replicate sample (Colle-15 replicate) from core segment 15 to examine the reproducibility of the results. Each sample contained between 230 and 870 g of ice. The samples were processed identically to the Jungfraujoch snow samples following the BRUGGER method, involving the same protocol modification for the second marker treatment (Fig. 1).

2.3. Pollen analysis

A pollen sum of 500 grains per sample was counted except for the high-alpine ice samples from Colle Gnifetti for which we reached a pollen sum of 350 due to low pollen concentrations. Pollen identification was conducted under a light microscope at 400 \times magnification following Beug (2004) and the reference collection of the Institute of Plant Sciences at University of Bern. Pollen percentage calculations are based on the terrestrial pollen sum, and concentrations were standardized to one liter of water. *Lycopodium* and *Eucalyptus* markers were counted alongside pollen and we obtained a minimum sum of 1000 marker grains for each sample.



* Modified for standardized comparison in this study according to flowchart in Fig. 1

Fig. 2. Marker ratio of *Lycopodium* (added prior to sample processing) to *Eucalyptus* (added before mounting in glycerine) for standard snow replicate samples from Jungfraujoch (Jung-1 to Jung-10). Sample preparation according to flowcharts in Figure 1 (modified for the standardized method comparison with two markers). Ideal marker relationship (dashed lines) with confidence intervals for marker tablet uncertainties based on standard deviations of tablet content (grey). Marker ratio and average deviation of markers from ideal marker ratio (% of 0.72) indicating average sample loss during processing with R^2 as a measure of correlation strength indicating loss variability.

2.4. Numerical analysis

We estimated average deviation of marker loss in percentages from the ideal marker ratio *Lycopodium* : *Eucalyptus* (0.72) based on the mean content of one marker tablet with one standard deviation of each marker to indicate the range of the ratio uncertainty for the tablets (0.66–0.78). We calculated coefficients of determination (R^2) for the marker ratios for each extraction method of the snow replicate samples and for the marker ratio of the high-alpine ice core samples as well as for pollen percentages of the two Colle Gnifetti replicate samples of segment Colle-15, with all pollen types >1%, including and excluding vesiculate pollen taxa. To test statistically potential differences between the effects of the extraction methods, we conducted one-way ANOVA followed by a Tukey-Kramer post-hoc test for the marker ratios and the vesiculate pollen percentages for all 60 snow samples grouped by extraction methods.

We performed ordination analyses to visualize the distance between the pollen assemblages of the 60 snow samples. The short length of the first axis (1.48 Std dev. units) of a detrended correspondence analysis (DCA, by segments) for the pollen percentage dataset of the snow replicate samples justifies using linear ordination methods (ter Braak and Prentice, 1988). Principal component analysis (PCA, ter Braak and Šmilauer, 2002) was performed based on a correlation matrix for pollen percentages (all taxa >1%), which is the standard unit to present pollen data (Maher, 1981; Faegri and Iversen, 1989). Additionally, we conducted PCA with the same explanatory variable based on a covariance matrix for pollen concentrations (grains l^{-1}) standardized by norm to assess potential absolute differences of single pollen types independently from each other (data not shown). Ordinations were carried out using CANOCO version 4.5 (ter Braak and Šmilauer, 2002).

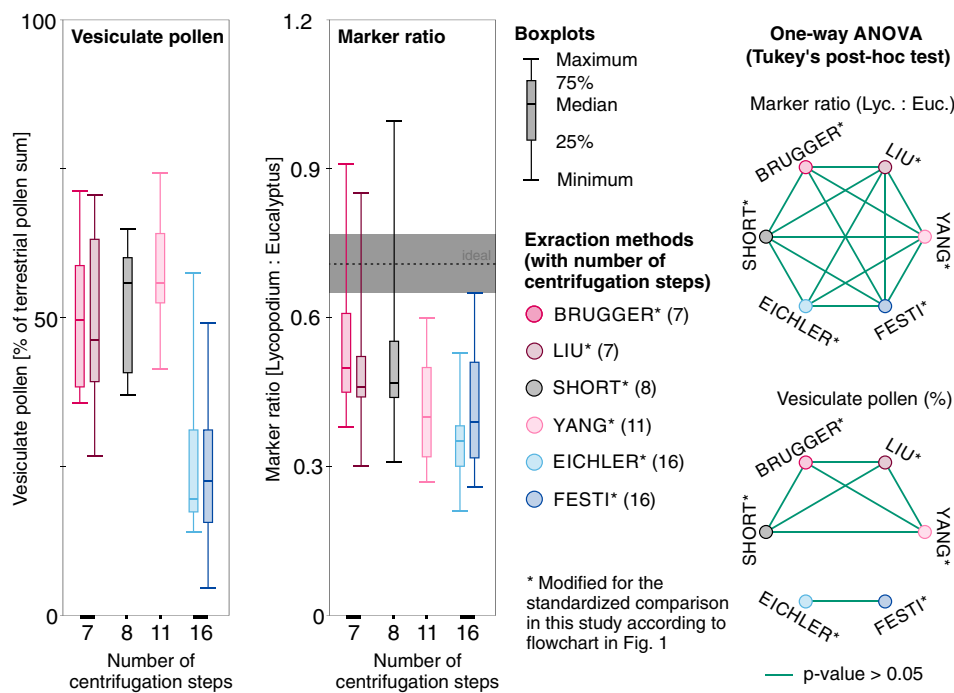


Fig. 3. Left: Boxplots for vesiculate pollen (as percentages of the terrestrial pollen sum, left side) and the counted marker ratio (*Lycopodium* : *Eucalyptus* with indication of the ideal marker ratio and one standard deviation uncertainty in grey, right side) for the different extraction methods sorted according to number of involved centrifugation steps including the physical and chemical treatment for 400 mL water (see numbers on the x-axis and in brackets in the method description) applied to standard snow replicate samples from Jungfraujoch. Right: One-way ANOVA followed by Tukey-Kramer post-hoc test results of marker ratios (top) and vesiculate pollen percentages among the different extraction methods. Circles joined by green lines indicate samples for which the tests did not provide evidence for statistical differences at the $p = 0.05$ level. All protocols modified from original lab protocols for the standardized comparison with a second marker (see flowchart in Fig. 1).

3. RESULTS AND INTERPRETATION

3.1. Marker tablet reliability

Given that the tablets completely dissolved in HCl, the added marker did not alter the quality of the samples. Instead, we assume that the marker spores and pollen increase the quality by adding more mass to the samples, thus contributing mass to and strengthening the organic pellet formed at the base of the centrifuge tubes (Moore and others, 1991). The tablets contained only the declared spores and pollen and the screening of the entire microscopic slides yielded no foreign materials (e.g. other pollen, charcoal) as potentially deriving from contamination during the tablet production process (see Festi and others, 2016). The ratio of *Lycopodium/Eucalyptus* grains was in all 10 samples of the marker reliability test within the expected ratio (0.66–0.78 *Lycopodium/Eucalyptus*) confirming that the marker tablets are reliable to use for our quantitative investigations.

3.2. Comparison of microfossil extraction methods with standard snow samples

All 60 samples yielded countable microfossil slides. The average marker ratio *Lycopodium/Eucalyptus* in the replicate snow samples processed with the BRUGGER method is 0.56 (22.1% average deviation from the ideal marker ratio i.e. 0.72). The number of counted *Lycopodium* spores is strongly correlated with the counted *Eucalyptus* pollen (R^2 of 0.94; Figs 2 and 3), suggesting that this pollen loss resulted in a systematic (and thus predictable) shift in the estimated pollen concentrations. Slightly higher marker deviation with 24% (average ratio = 0.55) is reached with the filtration-based SHORT method and 29.8% deviation (average ratio = 0.51)

with the related LIU method, while the R^2 of 0.87 reached in both methods is somewhat reduced compared with the BRUGGER method (R^2 of 0.94, Fig. 2). Extraction methods which involve many centrifugation steps without bottom freezing (EICHLER, FESTI, YANG) result in higher average marker deviation and lower R^2 (Figs 2 and 3, marker ratio = 0.36–0.42, 41–50% deviation, $R^2 = 0.81$ –0.86) suggesting that, on average, considerable numbers of *Lycopodium* spores (and thus of the targeted microfossils) are lost during water removal and chemical treatment and that the loss variability is larger with these extraction methods. However, ANOVA for the marker ratio differences between the extraction methods (Fig. 3) suggests that only the best performing BRUGGER and the least performing EICHLER method yield statistically significant differences in the mean marker ratios.

The pollen assemblages in all samples are dominated by *Picea abies*, *Pinus sylvestris*-type (both vesiculate = pollen with air bladders), *Alnus* and Poaceae with lower amounts of *Betula*, *Plantago lanceolata*-type, Cyperaceae and small pollen grains of *Castanea* and *Urtica* (Fig. 4). *Picea abies* and *Pinus sylvestris*-type pollen percentages and concentrations are generally higher in samples processed with evaporation- (BRUGGER, LIU, YANG) or filtration-based (SHORT) methods compared with samples processed with solely centrifugation-based methods (EICHLER, FESTI), suggesting that centrifugation may reduce vesiculate pollen grains compared with nonvesiculate pollen grains (Fig. 4). The large and relatively constant amount of *Urtica* (grain diameter ~10–15 μm , 15–20% abundance in all replicates from Jung-2) and *Alnus*, Poaceae and *Betula* (grain diameter 20–30 μm) in all replicate samples indicates that the six tested methods do not induce biases related to pollen grain size in the small to medium size range. Assessing differences in the amount of

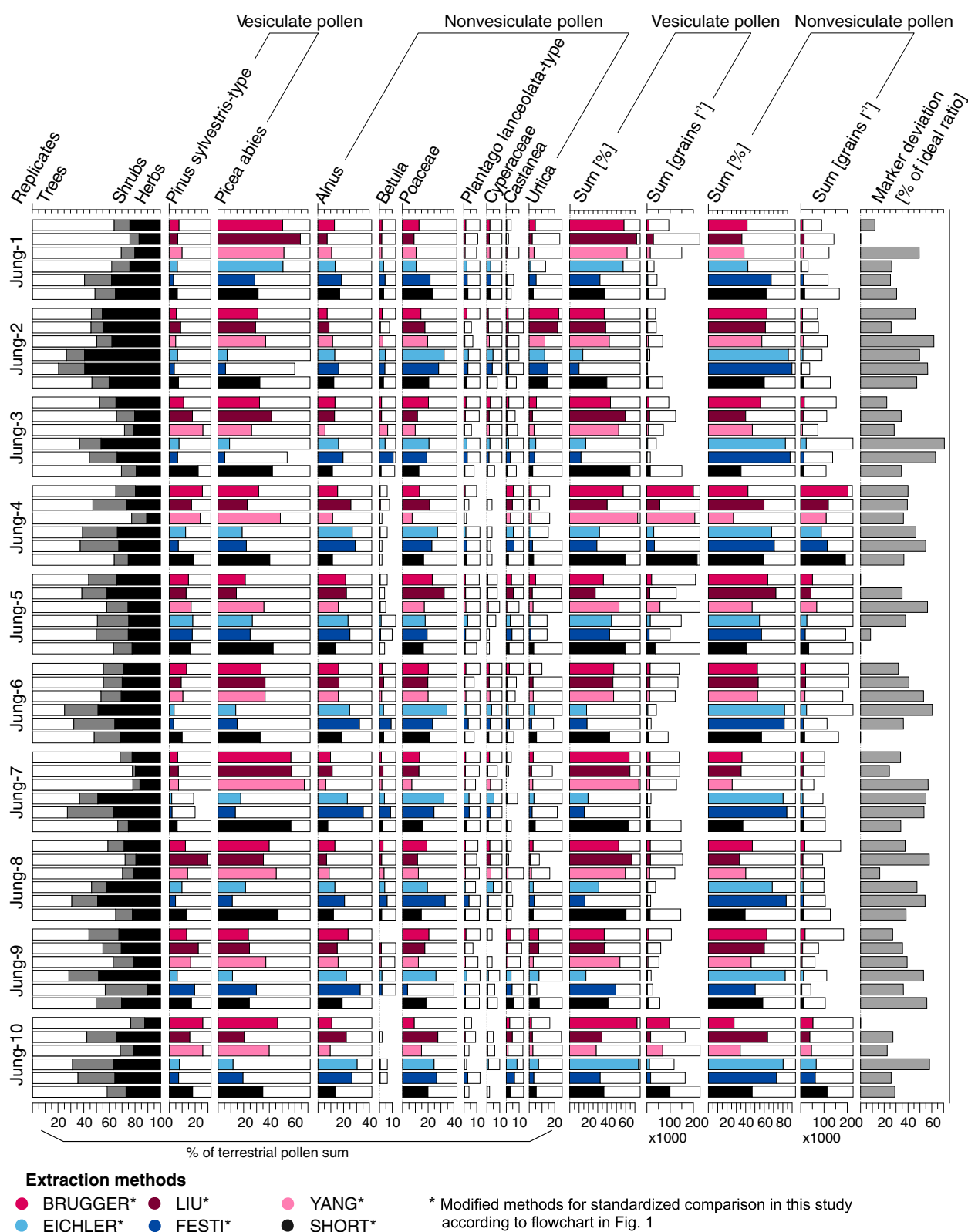


Fig. 4. Pollen percentage diagram of all pollen types more common than 5% in any one sample based on the terrestrial pollen sum and concentrations [grains l⁻¹] of vesiculate and nonvesiculate pollen for standard snow samples from Jungfrauoch. Each sample (Jung 1–10) was divided into replicate samples that were processed with six extraction methods (Modified from original protocols for a standardized comparison with a second marker, see flowchart in Fig. 1). Hollow bars = 10× exaggeration.

large nonvesiculate pollen (>50 µm, e.g. *Larix*) was not possible as these pollen types were too rare in our snow samples. The pollen concentrations in the snow replicate samples vary between 12000 and 65000 grains l⁻¹ (average 26000 grains l⁻¹) except samples Jung-4 and Jung-10 which contain five to 10 times higher pollen concentrations (Figs 3

and 4). Nonvesiculate pollen shows only small variability among snow replicate samples while vesiculate pollen concentrations show a much larger variability confirming that vesiculate pollen is mainly responsible for the differences in the percentages of the replicate samples, and therefore also of the main pollen percentage diagram for trees, shrubs and herbs.

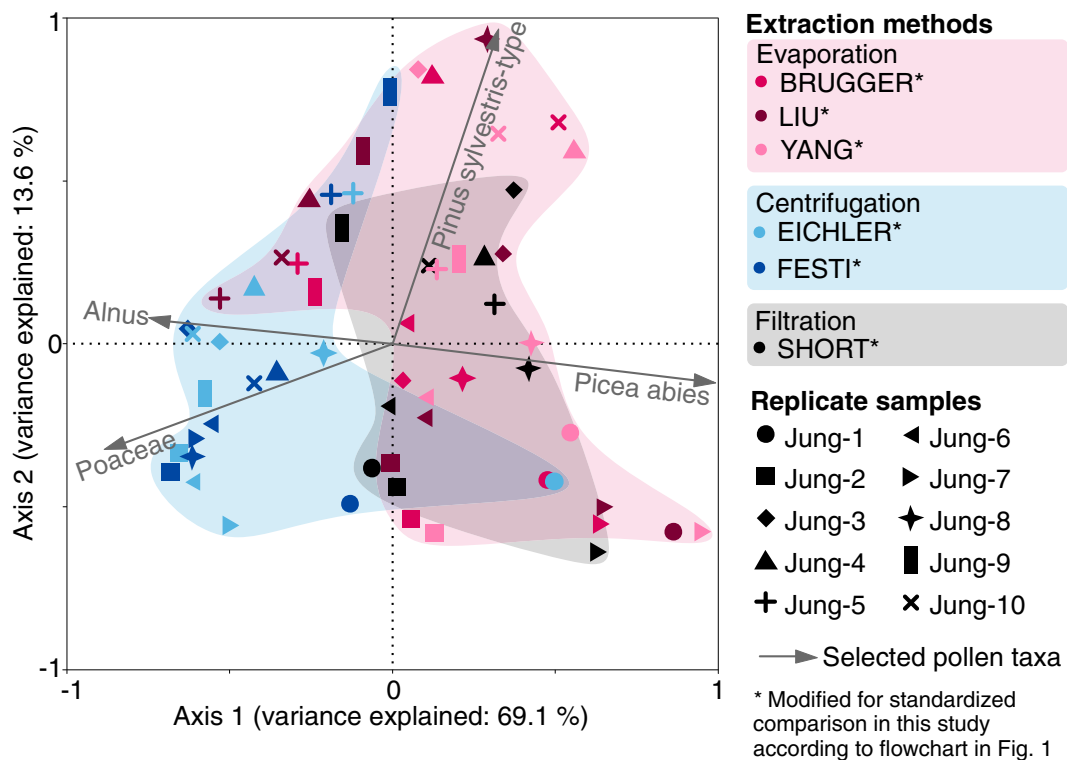


Fig. 5. Principal component analysis of pollen assemblages (percentages of the terrestrial pollen sum) for standard snow replicate samples from Jungfraujoch processed with six different microfossil extraction methods modified from original lab protocols for a standardized comparison with a second marker; see flowchart in Fig. 1. Colors refer to extraction methods grouped according to the main water elimination procedures: evaporation, centrifugation and filtration. Symbols refer to replicate samples.

The first PCA axis for the pollen percentages of the snow replicate samples explains 69.1% of the variance, and separates samples that used evaporation (BRUGGER, LIU and YANG) and filtration (SHORT) from samples that underwent many centrifugation steps without bottom freezing (EICHLER and FESTI methods). These latter samples contain generally lower amounts of *Picea abies* pollen (Fig. 5). The second axis explains 13.6% of the variance, and possibly reflects *Pinus sylvestris*-type abundance in the samples. Thus, the high cumulative variance explained by the two axes (82.7%) suggests that the vesiculate pollen abundance in the pollen assemblages may be strongly affected by the microfossil extraction procedure. Analyses with absolute values (pollen concentrations in grains l^{-1}) yield comparable results confirming the ordination results of the percentage dataset (data not shown).

The boxplots (Fig. 3) for vesiculate pollen percentages grouped by methods also indicate a selective vesiculate pollen loss with centrifugation-based methods without bottom freezing (EICHLER and FESTI by ~25%). It also seems that pollen composition among samples is characterized by a slightly higher variability compared with the evaporation and filtration-based methods. ANOVA for vesiculate pollen percentages grouped by extraction methods (Fig. 3) provides no evidence for statistically significant differences between samples processed with evaporation and filtration-based methods (BRUGGER, LIU, SHORT and YANG). For the centrifugation-based methods EICHLER and FESTI, ANOVA also provides no evidence for statistically significant differences in vesiculate pollen percentages. However, the analyses indicate statistically significant differences between these samples and those processed with the four other methods. Both ANOVA and visual examination of

the data, therefore, suggest a distinct difference between evaporation- or filtration-based methods and solely centrifugation-based methods in respect to the percentage of vesiculate pollen, with a clearly larger loss of vesiculate pollen in the EICHLER and FESTI methods. In contrast, the influence of different extraction methods on the marker ratio is less distinct (Fig. 3). This suggests a disproportionate effect of the solely centrifugation-based methods on vesiculate pollen (i.e. EICHLER and FESTI methods involving 16 centrifugation steps for 400 mL water, Fig. 3).

3.3. Validation of the BRUGGER method with high-alpine glacier ice samples

We applied the BRUGGER method to ice samples from the high-alpine Colle Gnifetti glacier because it yielded the smallest loss among all approaches used in our comparison with snow replicates from Jungfraujoch. The average marker ratio (0.57, 20.2% deviation from ideal ratio) in the high-alpine ice samples is very similar to the average marker ratio in the snow replicate samples processed with the same method (0.56, 22.1%), suggesting minor losses of *Lycopodium* and thus microfossils. The slightly lower correlation between *Lycopodium* spores and *Eucalyptus* pollen ($R^2 = 0.78$, Fig. 6a), when compared with snow samples (Fig. 6a), may be explained by properties inherent to these specific samples, for example low concentrations of insoluble organic and inorganic particles leading to smaller pellets during the concentration process. Indeed the microscopic slides from ice contained fewer dust particles than those from snow.

Pollen concentrations in the high-alpine ice core samples from Colle Gnifetti are low when compared with the snow

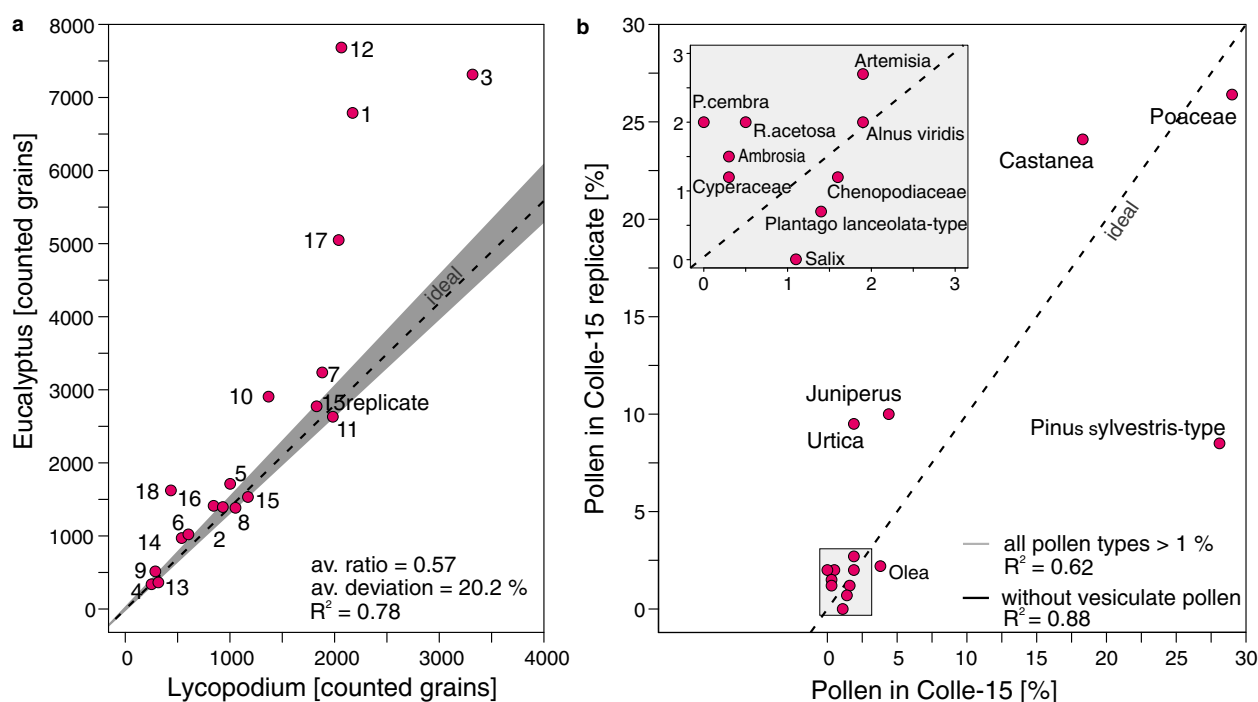


Fig. 6. Application of the BRUGGER method to Colle Gnifetti ice samples. Protocol with second marker application (see flowchart in Fig. 1). a) Marker ratio of *Lycopodium* spores (added prior to sample processing) and *Eucalyptus* pollen (added before mounting in glycerine) for 18 ice samples and one replicate of sample 15 from Colle Gnifetti glacier. Ideal marker relationship (9666 *Lycopodium* spores : 13500 *Eucalyptus* pollen = 0.72; dashed line) with tablet uncertainty (grey). b) Pollen assemblage comparison of sample 15 and its replicate (15 replicate) with all taxa presented as percentages of the terrestrial pollen sum. All pollen types > 1 % in one of the samples are shown. Dashed black line indicates ideal 1:1 pollen percentage ratio. *P. cembra* = *Pinus cembra*, *R. acetosa* = *Rumex acetosa*-type. Insert box: Magnification for pollen types with percentages between 1 and 3.

samples from Jungfraujoch, ranging between 310 and 18300 grains l^{-1} (mean of 6600 grains l^{-1}) vs. 12000–400000 grains l^{-1} (115000 grains l^{-1}), respectively. This finding may reflect the altitudinal difference of 1000 m between Colle Gnifetti (4450 m a.s.l.) and Jungfraujoch (3400 m a.s.l.), which markedly increases the distance to the vegetation at lower altitudes (colline to alpine belts between ~200 and 3000 m a.s.l.; Ellenberg, 1996) that produces the palynological signal. Pollen percentages for taxa > 1% in the two replicate samples of Colle-15 are very similar resulting in a high R^2 between pollen percentages of different taxa in these two samples (0.88 for pollen > 1% occurrence without vesiculate). The exception is *Pinus sylvestris*-type which occurs at 27% in Colle-15 and 10% in the Colle-15 replicate resulting in a much lower R^2 (0.62, Fig. 6b), if the vesiculate pollen taxa are included in the dataset (Fig. 6b). This suggests a high reproducibility of most pollen taxa (e.g. *Poaceae* and *Castanea*). The vesiculate morphology of *Pinus sylvestris*-type may influence the reproducibility of its abundance compared with other pollen taxa with a nonvesiculate morphology, while *Pinus cembra* values (2% vs. 0%, respectively) are too low to be assessed. Even though the statistical power of two high-alpine ice core replicates is limited, these results strongly support the outcome of the snow replicates as discussed in Section 3.2.

4. DISCUSSION

4.1. The necessity of marker application

Adding microfossil markers to palynological sediment samples has been a standard in palynology since the early

1970s (e.g. Stockmarr, 1971; Moore and others, 1991; Dark and Allen, 2005; Finsinger and Tinner, 2005; Maher and others, 2012; Brugger and others, 2016; Campbell and others, 2016; Rey and others, 2017). This allows quantitative pollen concentration and influx estimates (Stockmarr, 1971; Birks and Gordon, 1985), which cannot be achieved otherwise unless the entire sample is counted (von Post, 1916; Welten, 1944; Moore and others, 1991). Adding markers also helps to estimate losses during pollen extraction. Some recent studies tend to avoid adding markers to the palynological samples because of the costs, the additional labor or presumed contamination issues (Festi and others, 2016). Based on our results presented here, we can reject these speculations about contamination issues when using commercially available, standardized and quality checked marker tablets (e.g. *Lycopodium* tablets provided by the University of Lund; Maher, 1981).

Our results clearly show that microfossil loss during pollen extraction from ice samples is inevitable (i.e. > 20% of the *Lycopodium* marker is lost). This loss occurs while initially reducing the water content (e.g. evaporation, filtration, centrifuging followed by decanting), during the chemical treatment involving inevitable centrifuging steps in all tested methods and after sample processing. The very low amount of pollen in ice and snow samples is not easy to extract from the centrifuge tubes, to be fixed without losses on the slide and partly becomes covered under the microscopic cover slip margins. These losses strongly affect any absolute microfossil counts (e.g. pollen, charcoal particles in ice core samples) that do not add markers. Thus, marker application is imperative to estimate realistic absolute values (concentrations, influx) and as a control for total sample loss

(Stockmarr, 1971; Peck, 1974; Finsinger and Tinner, 2005), particularly for ice samples.

4.2. Influence of specific ice sample properties on pollen extraction

Ice samples have specific properties that may explain differences in the microfossil behavior during laboratory processing compared with sedimentary samples. The microfossil deposition on glaciers is different than the deposition in lake and mire sediment archives. Fresh pollen, spores and other microfossils (e.g. charcoal) get directly covered with snow after deposition on the surface and incorporated into the ice. This is unlike pollen deposited in lake sediments, which only settles to the lake bottom when it is saturated with water and dense enough to sink (Faegri and Iversen, 1989; Dark and Allen, 2005). A special behavior of vesiculate pollen compared with nonvesiculate pollen was demonstrated in transect studies in lakes, with shore sediments enriched in vesiculate pollen compared with sediments from the lake center. This pattern is explained by the fact that vesiculate pollen floats for a long time and is thus transported to the shore where it accumulates (Ammann and Tobolski, 1983). The different behavior of vesiculate and nonvesiculate pollen was also observed in floating pollen traps in lakes (Giesecke and Fontana, 2008). Early controlled laboratory experiments on the pollen floating behavior confirmed that the majority of vesiculate pollen (e.g. *Pinus* sp.) floated on the water surface in still standing glass beakers for hours while most fresh nonvesiculate pollen (e.g. 95–100% *Corylus*, *Quercus*, *Juniperus* and *Larix*) sank within the first 5 minutes after the experiment started (Hopkins, 1950). We thus assume that pollen of vesiculate pollen floated on the water surface after thawing of the ice samples and was preferentially lost during the subsequent decanting of the centrifuge tubes. A major difference between ice and sediment samples is that while pollen is frozen in ice, sediment pollen is soaked in water over years to millennia, potentially reducing its floating capacity. The high floating capacity of well-preserved vesiculate pollen in ice may explain the larger variability of vesiculate compared with nonvesiculate pollen (Fig. 4). This finding is important because it implies a negative bias on vesiculate pollen concentration estimates from ice core records. Peck (1972), Davis and Brubaker (1973) and Holmes (1990) reported differences of settling time among nonvesiculate pollen depending on the size. However, based on our results we cannot confirm a larger variability of pollen grains with small diameters and lower densities compared with larger grains (e.g. *Urtica* and *Castanea* vs. *Alnus* and Poaceae).

While the morphology of vesiculate pollen is an evolutionary advantage for wind and water pollination (the latter because buoyancy aids floating upwards in a liquid drop into the ovules; Owens and others, 1998; Runions and others, 1999), it is a clear disadvantage for laboratory processing of palynological samples. This implies that thawed ice samples should stand around some days to extend the water saturation and sinking time for pollen before applying centrifugation steps. This waiting time is inherent to evaporation-based methods (BRUGGER; LIU; YANG) where the pollen stays in the water for several days during the initial evaporation. Similarly, filtration-based methods (SHORT; or the method in Nakazawa and others, 2005) circumvent the problem of floating pollen in the first water reducing step. However, centrifuging samples may

also help vesiculate pollen to sink from a water surface by filling the air bladders with water (Hopkins, 1950) implying that increasing the centrifugation time in each centrifuging step may help to increase the number of sunken vesiculate pollen.

Microfossils from ice samples are not incorporated in a matrix as is typical for sediment samples from lakes or mires. We assume that this causes higher and more variable pollen loss during the pollen extraction from ice samples, especially during decanting of centrifuge tubes. Indeed, pollen assemblage differences were smaller in sediment samples as evidenced by a tentative comparison with replicate sediment samples from Moossee, Switzerland (unpublished results). Similarly, snow replicate samples from Jungfraujoch with most likely higher dust concentrations have a lower pollen loss variability than the high-alpine ice samples from Colle Gnifetti glacier, presumably since they form a more stable pellet at the bottom of centrifugation tubes after centrifugation. Our solution to this problem is to minimize pollen losses in ice samples by mimicking a matrix through shock-freezing of the tube bottom always after centrifuging and before decanting. The pollen quality was not affected by the shock-freezing procedure. However, our observations suggest that the low concentrations of suspended particles in the high-alpine ice core samples compared with samples with a sediment matrix may influence the sinking behavior during centrifugation. Comparing samples that are processed with identical laboratory protocols except for an alcohol treatment before centrifuging to lower the surface tension (EICHLER and FESTI methods) points to smaller pollen loss with an alcohol treatment (FESTI), although marker ratios are not significantly lower according to the comparison statistics (Fig. 3).

Our data indicate that losses increase with the number of increasing centrifuging steps (Fig. 3) suggesting that the number of centrifuging steps is crucial for microfossil losses. Remote archives at high altitudes (e.g. high alpine glaciers in the Andes above 6000 m a.s.l.; Liu and others) or high latitudes (e.g. Arctic sites, Short and Holdsworth, 1985; Hicks and Isaksson, 2006) usually demand high ice volumes (e.g. >400 mL) because of low pollen concentrations. Microfossil concentrations may also be more diluted in archives with high snow accumulation rates common in some mountain ranges (e.g. Neff and others, 2012; Schwikowski and others, 2013; Mariani and others, 2014). Given that the required sample volume for reliable analysis is much larger (i.e. several liters) in such extreme archives, evaporation methods appear best suited to reduce undesired pollen losses.

5. CONCLUSIONS

The BRUGGER extraction method is developed from existing evaporation-based water reducing methods (e.g. LIU protocol; Liu and others, 1998) with a newly invented step of freezing the tube bottom after centrifugation during the chemical treatment. Based on our results, the BRUGGER protocol can minimize microfossil loss if compared with the other approaches. However, the differences are statistically not pronounced. Our study highlights that pollen assemblages from ice cores processed with different microfossil extraction protocols challenge the reproducibility between records from different study sites. This is especially valid for sites where vesiculate pollen grains from conifers are an important component of pollen assemblages. Remote ice archives at extreme altitudes, high latitudes, or with high snow accumulation rates and consequently extremely low microfossil concentrations, need a special focus when processing samples for

palynological analysis. We show that significant losses are inevitable during processing of samples. Therefore, applying high-quality marker tablets prior to microfossil extraction from ice records is crucial if the goal is to produce absolute counts and not only percentages. Speculations about marker contaminations or other negative effects on the pollen assemblage quality can definitely be rejected. To conclude, we recommend following a strictly standardized protocol that includes high-quality marker tablets to obtain reliable concentration and influx estimates. Applying the new BRUGGER approach may contribute to minimizing microfossil losses and gaining reproducible results.

SUPPLEMENTARY MATERIAL

The supplementary material for this article can be found at <https://doi.org/10.1017/jog.2018.31>

ACKNOWLEDGEMENTS

We are grateful to Ruth Drescher-Schneider for providing *Eucalyptus* tablets, to Florencia Oberli for technical assistance with developing the new extraction method, to Daniela Festi, Klaus Oeggel and their colleagues from Innsbruck University for stimulating discussions, to Dimitri Osmont and Denis Alija for their help with sampling in the freezing room and at the Jungfrauoch station. We thank H.J.B. Birks and an anonymous reviewer for their constructive remarks, which substantially improved the manuscript. We acknowledge the SINERGIA project *Paleo fires from high-alpine ice cores* funded by the Swiss National Science Foundation (SNF grant 154450).

AUTHORS CONTRIBUTIONS

SO Brugger and E Gobet contributed equally.

REFERENCES

- Ambach W, Bortenschlager S and Eisner H (1966) Pollen-analysis investigation of a 20 m. Firn Pit on the Kesselwandferner (Ötztal Alps). *J. Glaciol.*, **6**(44), 233–236 (doi: 10.3189/S0022143000019249)
- Ammann B and Tobolski K (1983) Vegetational development during the late-Würm at Lobsigensee (Swiss Plateau) (studies in the late quaternary at Lobsigensee 1). *Rev. Paleobiol.*, **2**, 163–180
- Andreev AA, Nikolaev VI, Bolshiyaynov DY and Petrov VN (1997) Pollen and isotope investigations of an ice core from Vavilov ice cap, October Revolution Island, Severnaya Zemlya Archipelago, Russia. *Geogr. Phys. Quat.*, **51**(3), 379–389 (doi: 10.7202/033137ar)
- Benninghoff WS (1962) Calculation of pollen and spore density in sediments by addition of exotic pollen in known quantities. *Pollen Spores.*, **4**, 332–333
- Beug H-J (2004) *Leitfaden der Pollenbestimmung für Mitteleuropa und angrenzende Gebiete*. Pfeil, München
- Birks HJB and Birks HH (1980) *Quaternary palaeoecology*. Edward Arnold, London
- Birks HJB and Gordon AD (1985) *Numerical methods in quaternary pollen analysis*. Academic Press, London
- Bourgeois JC (1990) A modern pollen spectrum from Dye 3, south Greenland ice sheet. *J. Glaciol.*, **36**(124), 340–342 (doi: 10.3189/002214390793701363)
- Bourgeois JC (2000) Seasonal and interannual pollen variability in snow layers of Arctic ice caps. *Rev. Palaeobot. Palynol.*, **108**, 17–36 (doi: 10.1016/S0034-6667(99)00031-7)
- Bourgeois JC, Koerner RM, Gajewski K and Fisher DA (2000) A holocene ice-core pollen record from Ellesmere Island, Nunavut, Canada. *Quat. Res.*, **54**, 275–283 (doi: 10.1006/qres.2000.2156)
- Bourgeois JC, Gajewski K and Koerner RM (2001) Spatial patterns of pollen deposition in arctic snow. *J. Geophys. Res. Atmos.*, **106**(D6), 5255–5265 (doi: 10.1029/2000JD900708)
- Brugger SO and 13 others (2016) Long-term man–environment interactions in the Bolivian Amazon: 8000 years of vegetation dynamics. *Quat. Sci. Rev.*, **132**, 114–128 (doi: 10.1016/j.quascirev.2015.11.001)
- Campbell JFE, Fletcher WJ, Hughes PD and Shuttleworth EL (2016) A comparison of pollen extraction methods confirms dense-media separation as a reliable method of pollen preparation. *J. Quat. Sci.*, **31**(6), 631–640 (doi: 10.1002/jqs.2886)
- Dark P and Allen JRL (2005) Seasonal deposition of Holocene banded sediments in the Severn Estuary Levels (southwest Britain): palynological and sedimentological evidence. *Quat. Sci. Rev.*, **24**(1), 11–33 (doi: 10.1016/j.quascirev.2004.08.001)
- Davis MB and Brubaker LB (1973) Differential sedimentation of pollen grains in lakes. *Limnol. Oceanogr.*, **18**(4), 635–646 (doi: 10.4319/lo.1973.18.4.0635)
- Dietrich W (1923) *Die Erscheinungen an den Grenzflächen. In Einführung in die Physikalische Chemie für Biochemiker, Mediziner, Pharmazeuten und Naturwissenschaftler*. Springer, Berlin Heidelberg, 1–105
- Eichler A and 5 others (2011) An ice-core based history of Siberian forest fires since AD 1250. *Quat. Sci. Rev.*, **30**(9), 1027–1034 (doi: 10.1016/j.quascirev.2011.02.007)
- Ellenberg H (1996) *Vegetation Mitteleuropas mit den Alpen in ökologischer, dynamischer und historischer Sicht*. 5th edn. Ulmer, Stuttgart
- Fægri K and Iversen J (1989) *Textbook of pollen analysis*. 4th edn. Wiley, Winchester
- Festi D and 6 others (2015) A novel pollen-based method to detect seasonality in ice cores: a case study from the Ortles glacier, South Tyrol, Italy. *J. Glaciol.*, **61**(229), 815–824 (doi: 10.3189/2015JoG14J236)
- Festi D, Hoffmann DL and Luetscher M (2016) Pollen from accurately dated speleothems supports alpine glacier low-stands during the early Holocene. *Quat. Res.*, **86**(1), 45–53 (doi: 10.1016/j.yqres.2016.05.003)
- Feurdean A, Perşoiu A, Pazdur A and Onac BP (2011) Evaluating the palaeoecological potential of pollen recovered from ice in caves: a case study from Scărișoara Ice Cave, Romania. *Rev. Palaeobot. Palynol.*, **165**(1), 1–10 (doi: 10.1016/j.revpalbo.2011.01.007)
- Finsinger W and Tinner W (2005) Minimum count sums for charcoal concentration estimates in pollen slides: accuracy and potential errors. *Holocene*, **15**(2), 293–297 (doi: 10.1191/0959683605hl808rr)
- Fredskild B and Wagner P (1974) Pollen and fragments of plant tissue in core samples from the Greenland ice cap. *Boreas*, **3**(3), 105–108 (doi: 10.1111/j.1502-3885.1974.tb00668.x)
- Giesecke T and Fontana SL (2008) Revisiting pollen accumulation rates from Swedish lake sediments. *Holocene*, **18**(2), 293–305 (doi: 10.1177/0959683607086767)
- Herren PA and 6 others (2013) The onset of Neoglaciation 6000 years ago in western Mongolia revealed by an ice core from the Tsambagarav mountain range. *Quat. Sci. Rev.*, **69**, 59–68 (doi: 10.1016/j.quascirev.2013.02.025)
- Hicks S and Isaksson E (2006) Assessing source areas of pollutants from studies of fly ash, charcoal, and pollen from Svalbard snow and ice. *J. Geophys. Res. Atmos.*, **111**(D2), D02113 (doi: 10.1029/2005JD006167)
- Holmes PL (1990) Differential transport of spores and pollen: a laboratory study. *Rev. Palaeobot. Palynol.*, **64**(1–4), 289–296 (doi: 10.1016/0034-6667(90)90144-8)
- Hopkins JS (1950) Differential flotation and deposition of coniferous and deciduous tree pollen. *Ecology*, **31**(4), 633–641 (doi: 10.2307/1931580)
- Jenk TM and 9 others (2009) A novel radiocarbon dating technique applied to an ice core from the Alps indicating late Pleistocene ages. *J. Geophys. Res. Atmos.*, **114**(D14), D14305 (doi: 10.1029/2009JD011860)

- Koerner RM, Bourgeois JC and Fisher DA (1988) Pollen analysis and discussion of time-scales in Canadian ice cores. *Ann. Glaciol.*, **10**, 85–91 (doi: 10.1017/S0260305500004225)
- Konrad H, Bohleber P, Wagenbach D, Vincent C and Eisen O (2013) Determining the age distribution of Colle Gnifetti, Monte Rosa, Swiss Alps, by combining ice cores, ground-penetrating radar and a simple flow model. *J. Glaciol.*, **59**(213), 179–189 (doi: 10.3189/2013JoG12J072)
- Lang G (1994) *Quartäre Vegetationsgeschichte Europas: Methoden und Ergebnisse*. Gustav Fischer, Jena
- Lauber K and Wagner G (2012) *Flora helvetica*. 5th edn. Haupt, Bern
- Liu KB, Yao Z and Thompson LG (1998) A pollen record of Holocene climatic changes from the Dunde ice cap, Qinghai-Tibetan Plateau. *Geology*, **26**(2), 135–138 (doi: 10.1130/0091-7613(1998)026<0135:APROHC>2.3.CO;2)
- Liu KB, Reese CA and Thompson LG (2005) Ice-core pollen record of climatic changes in the central Andes during the last 400 yr. *Quat. Res.*, **64**(2), 272–278 (doi: 10.1016/j.yqres.2005.06.001)
- Liu KB, Reese CA and Thompson LG (2007) A potential pollen proxy for ENSO derived from the Sajama ice core. *Geophys. Res. Lett.*, **34**(9), L09504 (doi: 10.1029/2006GL029018)
- Maher LJ (1981) Statistics for microfossil concentration measurements employing samples spiked with marker grains. *Rev. Palaeobot. Palynol.*, **32**(2–3), 153–191 (doi: 10.1016/0034-6667(81)90002-6)
- Maher LJ, Heiri O and Lotter AF (2012) Assessment of uncertainties associated with palaeolimnological laboratory methods and microfossil analysis. In Birks H, Lotter A, Juggins S and Smol J, eds. *Tracking environmental change using lake sediments*. Springer, Dordrecht, 143–166 (doi: 10.1007/978-94-007-2745-8_6)
- Mariani I and 6 others (2014) Temperature and precipitation signal in two Alpine ice cores over the period 1961–2001. *Clim. Past*, **10**(3), 1093 (doi: 10.5194/cp-10-1093-2014)
- McAndrews JH (1984) Pollen analysis of the 1973 ice core from Devon Island Glacier, Canada. *Quat. Res.*, **22**(1), 68–76 (doi: 10.1016/0033-5894(84)90007-3)
- Moore PD, Webb JA and Collison ME (1991) *Pollen analysis*. Blackwell Scientific Publications, Oxford
- Nakazawa F and Fujita K (2006) Use of ice cores from glaciers with melting for reconstructing mean summer temperature variations. *Ann. Glaciol.*, **43**(1), 167–171 (doi: 10.3189/172756406781812302)
- Nakazawa F, Fujita K, Uetake J, Kohno M, Fujiki T, Arkhipov SM, Kameda T, Suzuki K and Fujii Y (2004) Application of pollen analysis to dating of ice cores from lower-latitude glaciers. *J. Geophys. Res. Earth. Surf.*, **109**(F4), F04001 (doi: 10.1029/2004JF000125)
- Nakazawa F and 6 others (2005) Dating of seasonal snow/firn accumulation layers using pollen analysis. *J. Glaciol.*, **51**(174), 483–490 (doi: 10.3189/172756505781829179)
- Nakazawa F and 7 others (2011) Establishing the timing of chemical deposition events on Belukha glacier, Altai Mountains, Russia, using pollen analysis. *Arct. Antar. Alp. Res.*, **43**(1), 66–72 (doi: 10.1657/1938-4246-43.1.66)
- Neff PD and 5 others (2012) Ice-core net snow accumulation and seasonal snow chemistry at a temperate-glacier site: Mount Waddington, southwest British Columbia, Canada. *J. Glaciol.*, **58**(212), 1165–1175 (doi: 10.3189/2012JoG12J078)
- Olivier S, and 8 others (2006) Temporal variations of mineral dust, biogenic tracers, and anthropogenic species during the past two centuries from Belukha ice core, Siberian Altai. *J. Geophys. Res. Atmos.*, **111**(D5), D05309 (doi: 10.1029/2005JD005830)
- Owens JN, Takaso T and Runions CJ (1998) Pollination in conifers. *Trends Plant Sci.*, **3**(12), 479–F04485 (doi: 10.1016/S1360-1385(98)01337-5)
- Papina T and 5 others (2013) Biological proxies recorded in a Belukha ice core, Russian Altai. *Clim. Past*, **9**(5), 2399–2411 (doi: 10.5194/cp-9-2399-2013)
- Peck RM (1972) Efficiency tests on the Tauber trap used as a pollen sampler in turbulent water flow. *New Phytol.*, **71**(1), 187–198 (doi: 10.1111/j.1469-8137.1972.tb04827.x)
- Peck RM (1974) A comparison of four absolute pollen preparation techniques. *New Phytol.*, **73**(3), 567–587 (doi: 10.1111/j.1469-8137.1974.tb02133.x)
- Preunkert S, Wagenbach D and Legrand M (2003) A seasonally resolved alpine ice core record of nitrate: comparison with anthropogenic inventories and estimation of preindustrial emissions of NO in Europe. *J. Geophys. Res. Atmos.*, **108**(D21), 10 (doi: 10.1029/2003JD003475)
- Reese CA and Liu KB (2002) Pollen dispersal and deposition on the Quelccaya Ice Cap, Peru. *Phys. Geogr.*, **23**(1), 44–58 (doi: 10.2747/0272-3646.23.1.44)
- Reese CA and Liu KB (2005) Interannual variability in pollen dispersal and deposition on the tropical Quelccaya Ice Cap. *Prof. Geogr.*, **57**(2), 185–197 (doi: 10.1111/j.0033-0124.2005.00471.x)
- Reese CA, Liu KB and Mountain KR (2003) Pollen dispersal and deposition on the ice cap of Volcán Paríacota, southwestern Bolivia. *Arct. Antar. Alp. Res.*, **35**(4), 469–474
- Rey F and 10 others (2017) Vegetational and agricultural dynamics at Burgäschisee (Swiss Plateau) recorded for 18,700 years by multiproxy evidence from partly varved sediments. *Veget. Hist. Archaeobot.*, **26**(6), 571–586 (doi: 10.1007/s00334-017-0635-x)
- Runions CJ, Rensing KH, Takaso T and Owens JN (1999) Pollination of *Picea orientalis* (Pinaceae): saccus morphology governs pollen buoyancy. *Am. J. Bot.*, **86**(2), 190–197 (doi: 10.2307/2656936)
- Santibañez P and 8 others (2008) Glacier mass balance interpreted from biological analysis of firn cores in the Chilean lake district. *J. Glaciol.*, **54**(186), 452–462 (doi: 10.3189/002214308785837101)
- Schwikowski M, Schläppi M, Santibañez P, Rivera A and Casassa G (2013) Net accumulation rates derived from ice core stable isotope records of Pío XI glacier, Southern Patagonia Icefield. *Cryosphere*, **7**(5), 1635–1644 (doi: 10.5194/tc-7-1635-2013)
- Short SK and Holdsworth G (1985) Pollen, oxygen isotope content and seasonally in an ice core from the penny Ice Cap, Baffin Island. *Arctic*, **38**(3), 214–218 (doi: 10.14430/arctic2136)
- Sigl M, and 10 others (2009) Towards radiocarbon dating of ice cores. *J. Glaciol.*, **55**(194), 985–996 (doi: 10.3189/002214309790794922)
- Stockmarr J (1971) Tablets with spores used in absolute pollen analysis. *Pollen Spores*, **XIII**(4), 615–621
- Ter Braak CJF and Prentice IC (1988) A theory of gradient analysis. *Adv Ecol Res*, **18**, 271–317 (doi: 10.1016/S0065-2504(03)34003-6)
- Ter Braak CJF and Šmilauer P (2002) *Canoco for windows version 4.5*. Biometris–Plant Research International, Wageningen
- Uetake J and 7 others (2006) Biological ice-core analysis of Sofiyskiy glacier in the Russian Altai. *Ann. Glaciol.*, **43**(1), 70–78 (doi: 10.3189/172756406781811925)
- Vareschi V (1934) Pollenanalysen aus Gletschereis. *Ber. Geobot. Inst. Eidgenöss. Tech. Hochsch. Stift. Rübel*, **1934**, 81–99
- von Post L (1916) Einige südschwedischen Quellmoore. *Bull. Geol. Inst. Univ. Upsala*, **15**, 219–278
- Welten M (1944) *Pollenanalytische, stratigraphische und geochronologische Untersuchungen aus dem Faulenseemoos bei Spiez* (Vol. 21). Huber, Zürich
- Yang B, Tang L, Bräuning A, Davis ME, Shao J and Jingjing L (2008) Summer temperature reconstruction on the central Tibetan Plateau during 1860–2002 derived from annually resolved ice core pollen. *J. Geophys. Res. Atmos.*, **113**(D24), D24102 (doi: 10.1029/2008JD010142)